

Modèles murins de la pré-éclampsie :  
des visions différentes et complémentaires de la pathologie humaine  
Murine models of preeclampsia:  
bringing a different and complementary vision to the human disease

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**37<sup>es</sup> JHTA**  
JOURNÉES DE L'HYPERTENSION ARTÉRIELLE  
DU DIAGNOSTIC AUX COMPLICATIONS

**14-15 décembre 2017**  
**Paris**

[www.jhta2017.fr](http://www.jhta2017.fr)

**11<sup>th</sup> INTERNATIONAL MEETING OF THE FRENCH SOCIETY OF HYPERTENSION**

Cité Universitaire (CIUP)

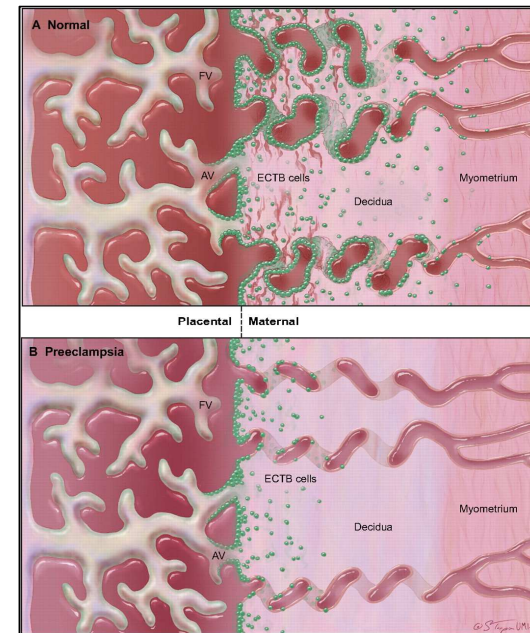
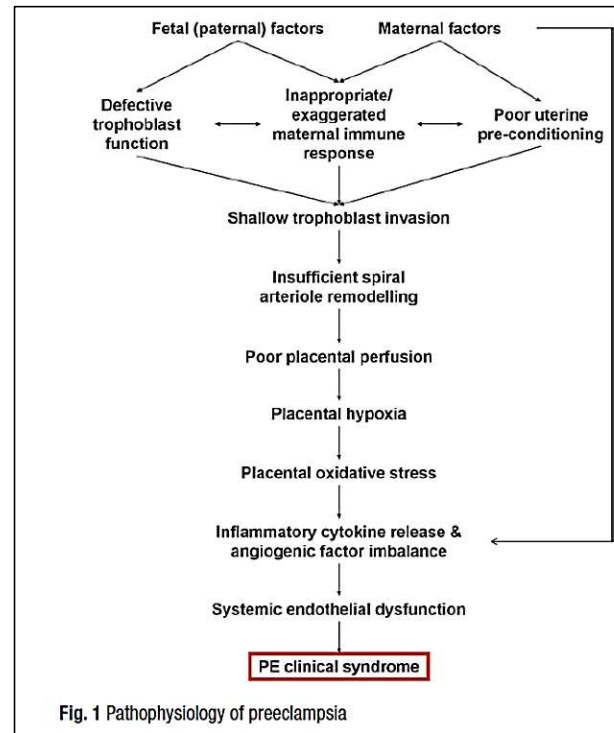
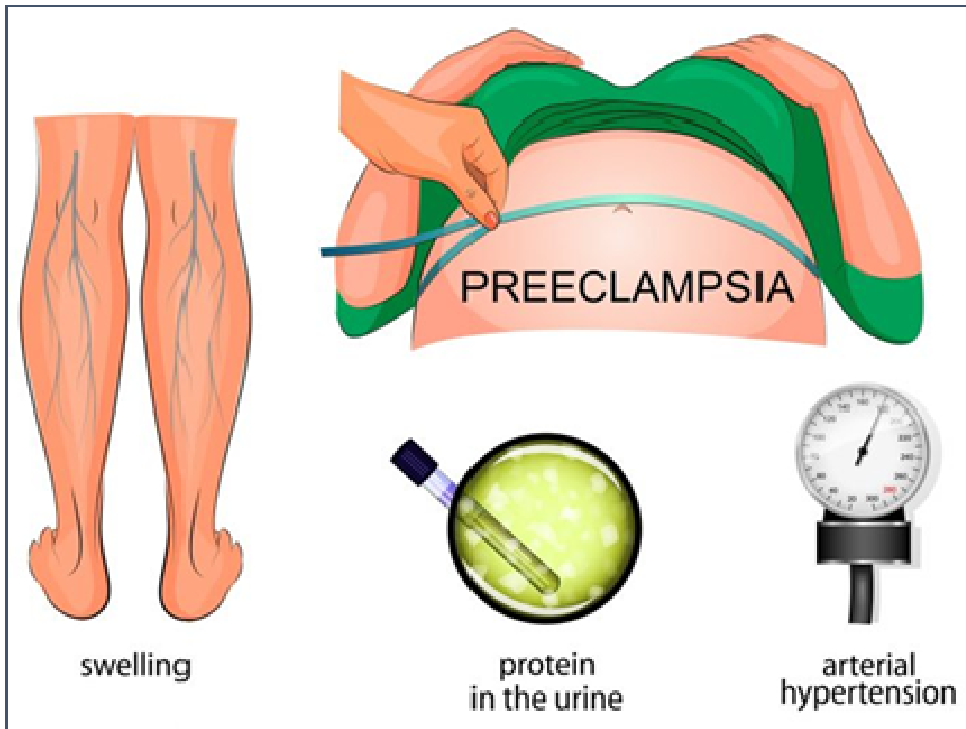
Société Française d'Hypertension Artérielle  
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Société Belge d'Hypertension

Société Française d'Hypertension Artérielle

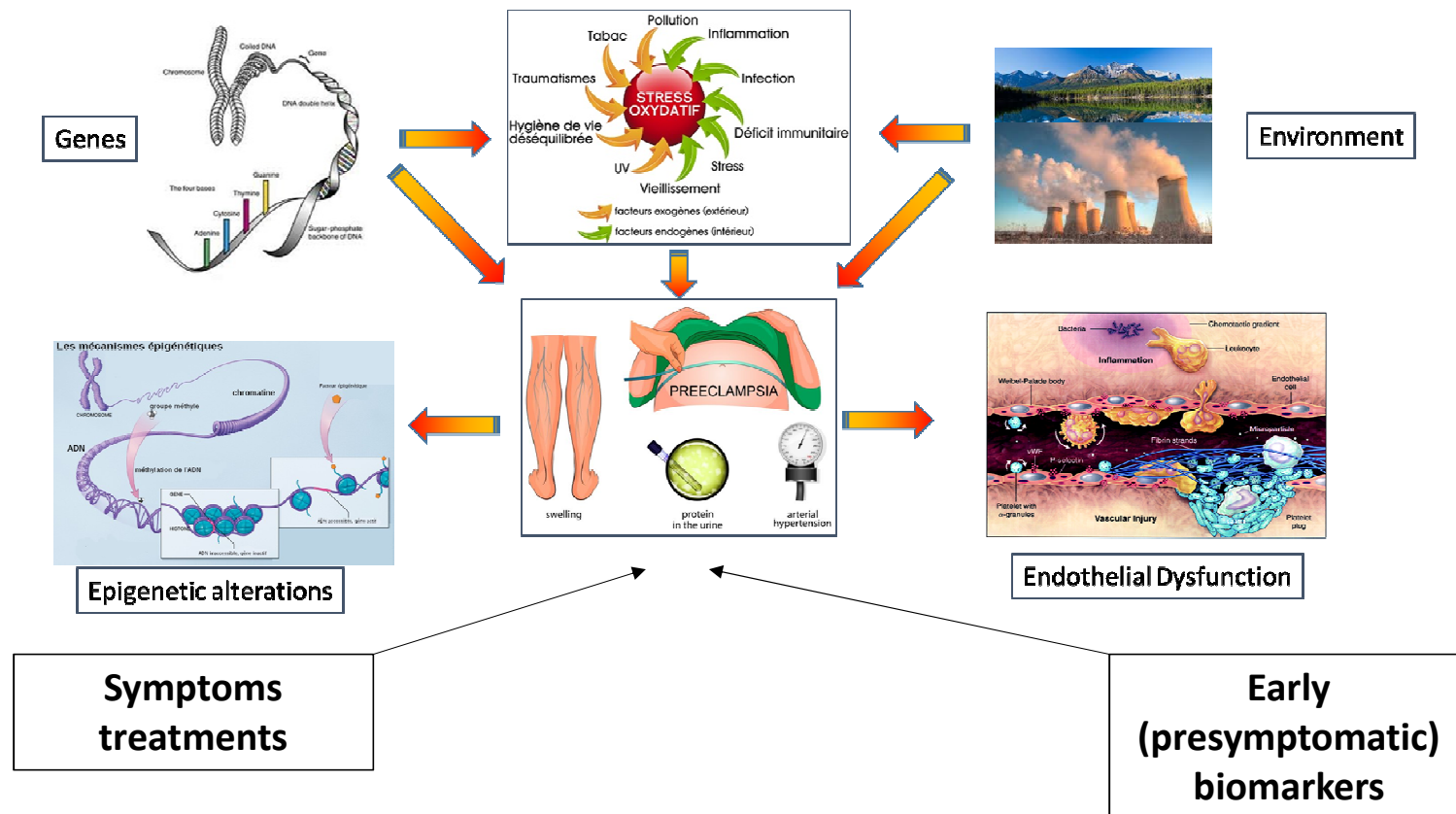
Société Suisse d'Hypertension

# Physiopathology of Preeclampsia



Padma Murthi and Cathy Vaillancourt (eds.), *Preeclampsia: Methods and Protocols*, Methods in Molecular Biology, vol. 1710, [https://doi.org/10.1007/978-1-4939-7498-6\\_5](https://doi.org/10.1007/978-1-4939-7498-6_5), © Springer Science+Business Media LLC 2018  
 Pennington, K.A. *et al*, *Dis. Mod. Mech.*, 5: 9-18, 2012

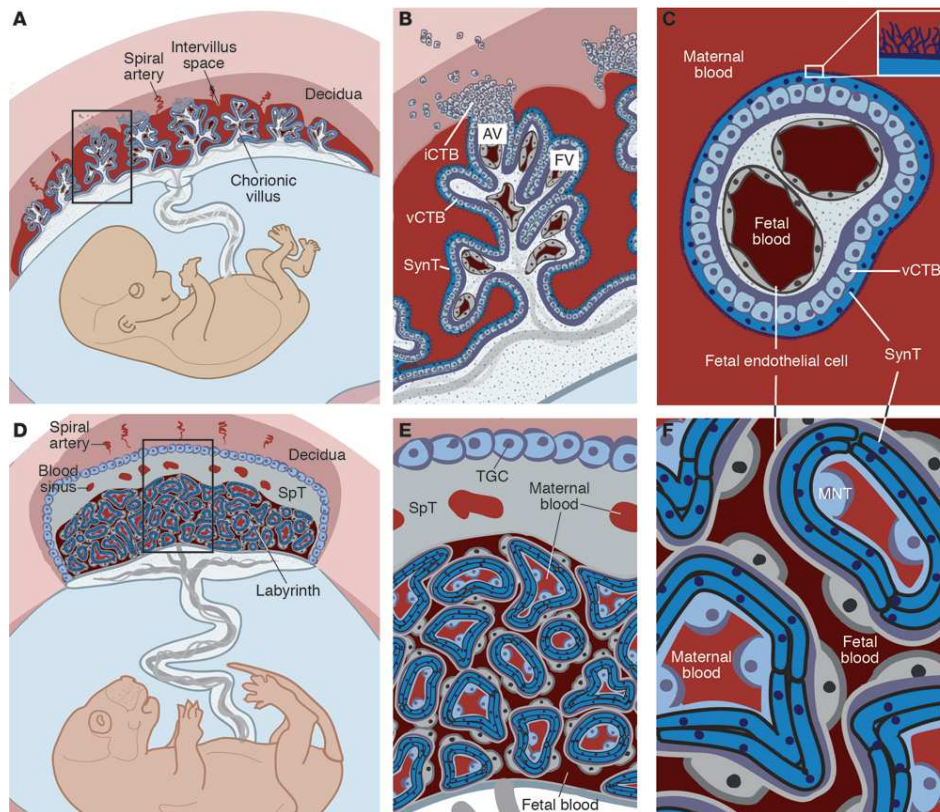
# Research questions in preeclampsia





# Mouse versus human placentation

- A discoid shape, hemochorial. Three cell layers separating maternal and fetal circulations.



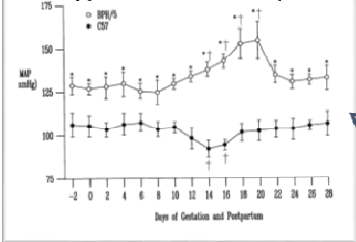
- The human placenta: monochorial, existence of placental villi in the intervillous space
- The murine placenta: Hemo-Trichorial, but the maternal blood accesses to the syncytiotrophoblast bilayer. Specific structure: junctional zone, labyrinth



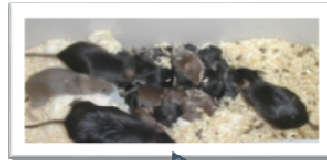
# Emulating preeclampsia in mice

## Genetics

Pre hypertensive mice (BPH/5)

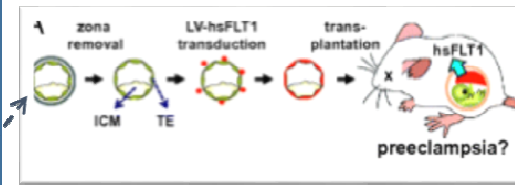


Crosses (CBAxDBA)

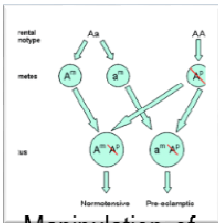
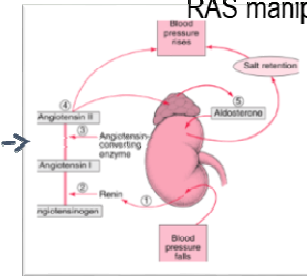


## Angiogenesis and Hypertension

Sflt1 overexpression



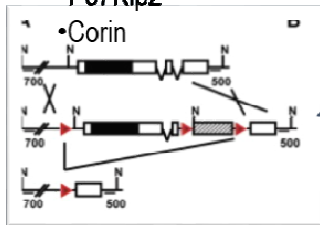
RAS manipulation



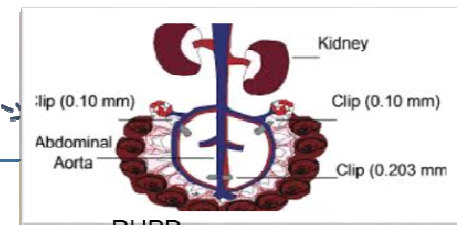
Manipulation of imprinted genes

Specific KO mice

- Comt
- P57Kip2
- Corin

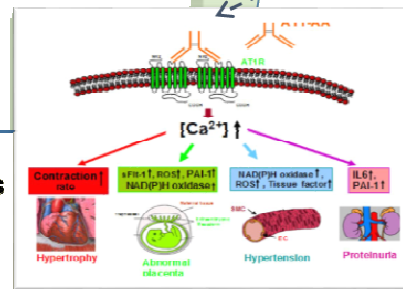


## Surgery

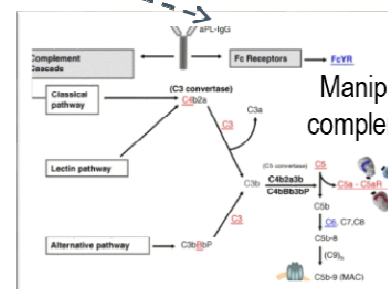


RUPP surgery

Injection of autoantibodies



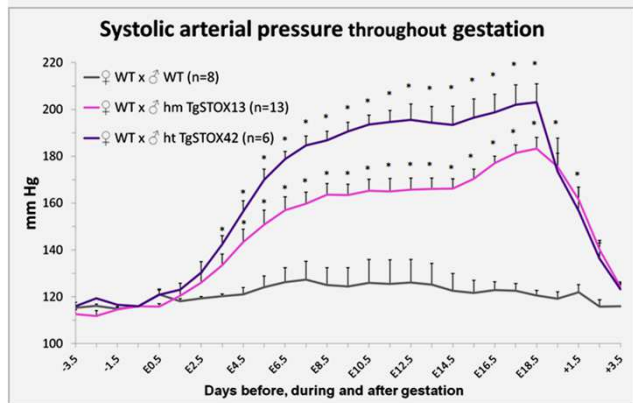
Manipulation of the complement cascade



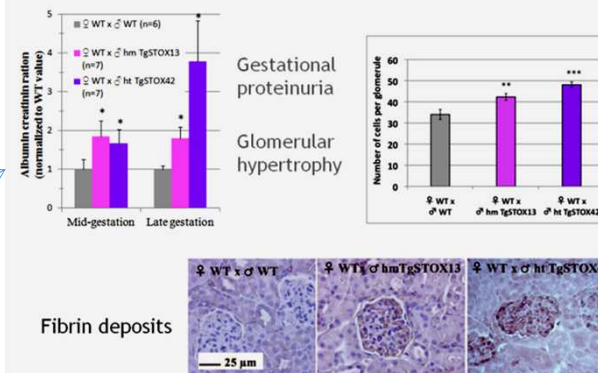
## Immunity and inflammation

# Foeto-placental expression of *STOX1* induces preeclampsia in mice

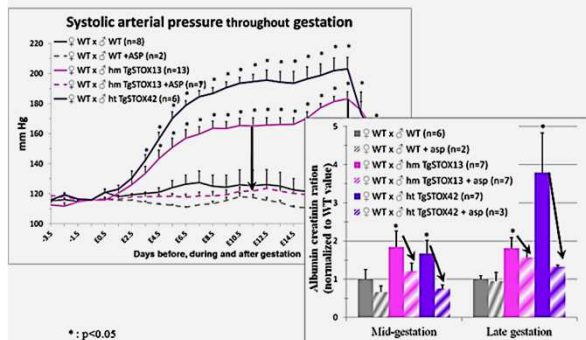
## PREGNANCY HYPERTENSION



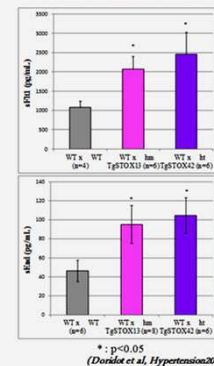
## PROTEINURIA AND KIDNEY HISTOLOGY



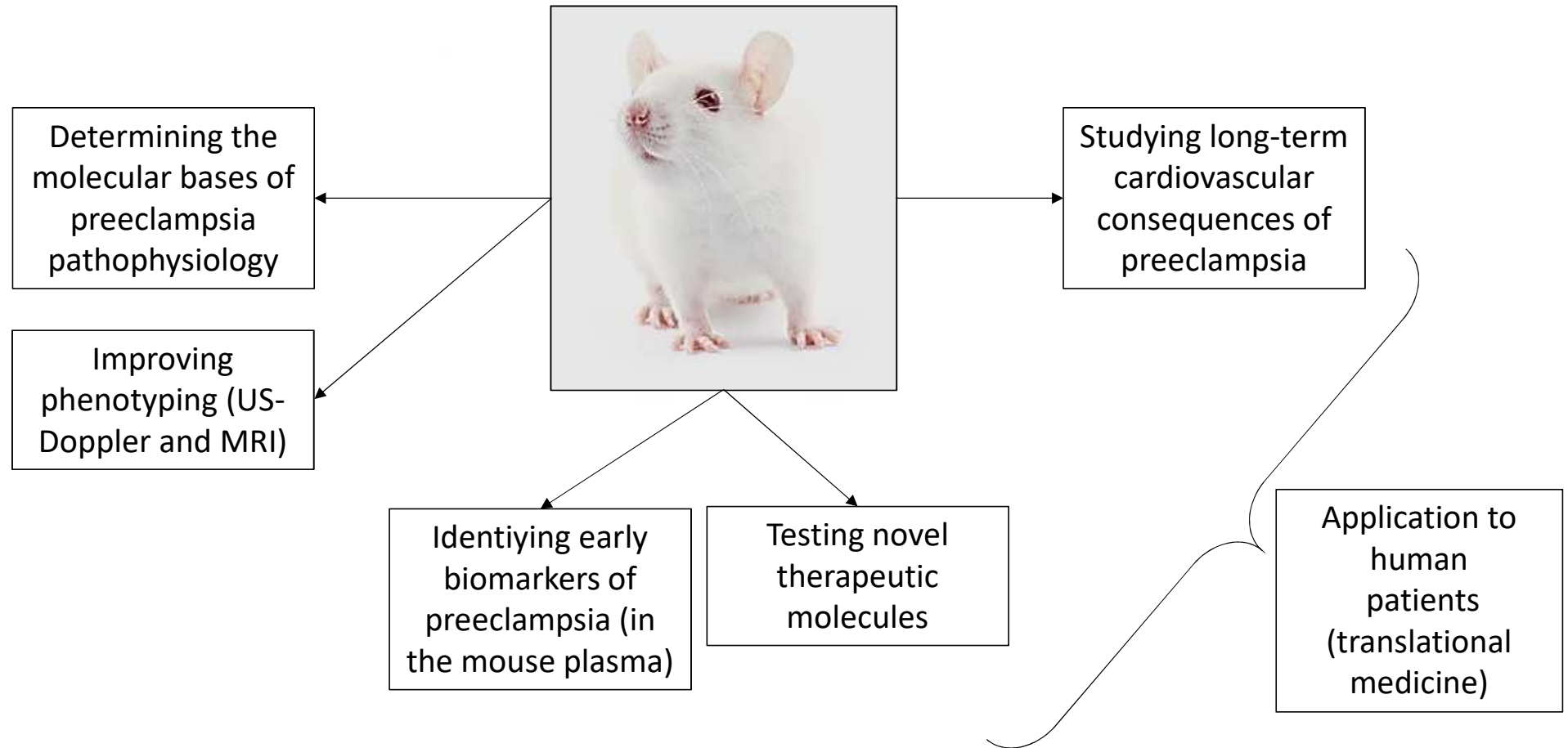
## ASPIRIN IMPACT



## ANTI-ANGIOGENIC FACTORS

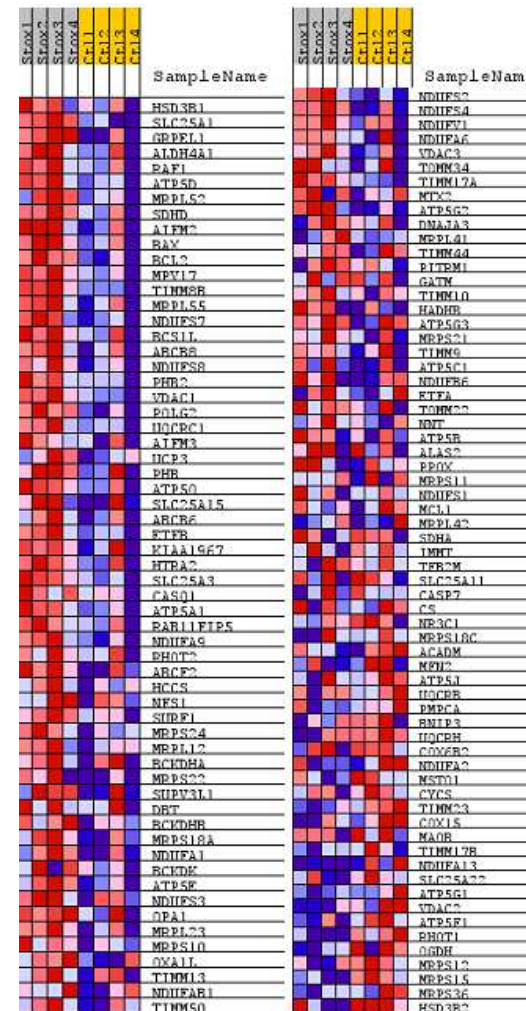
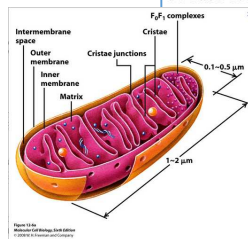
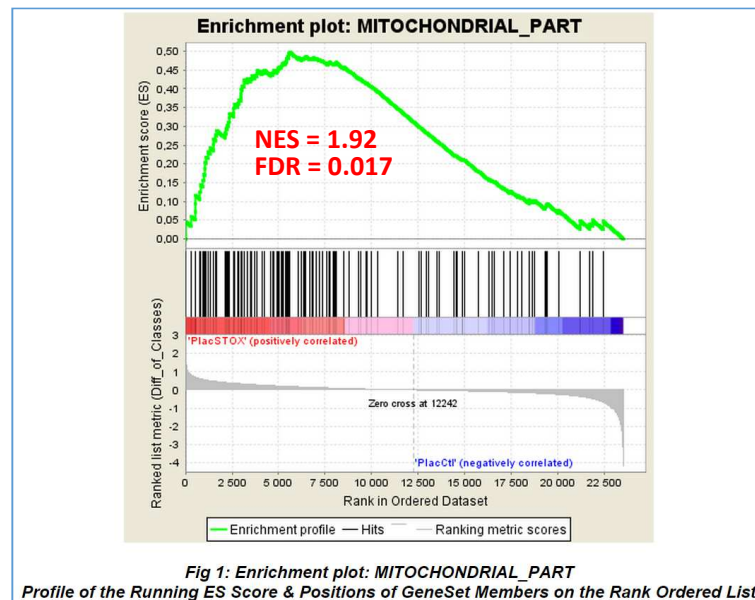


# Novel ongoing projects using preeclamptic STOX1 mice





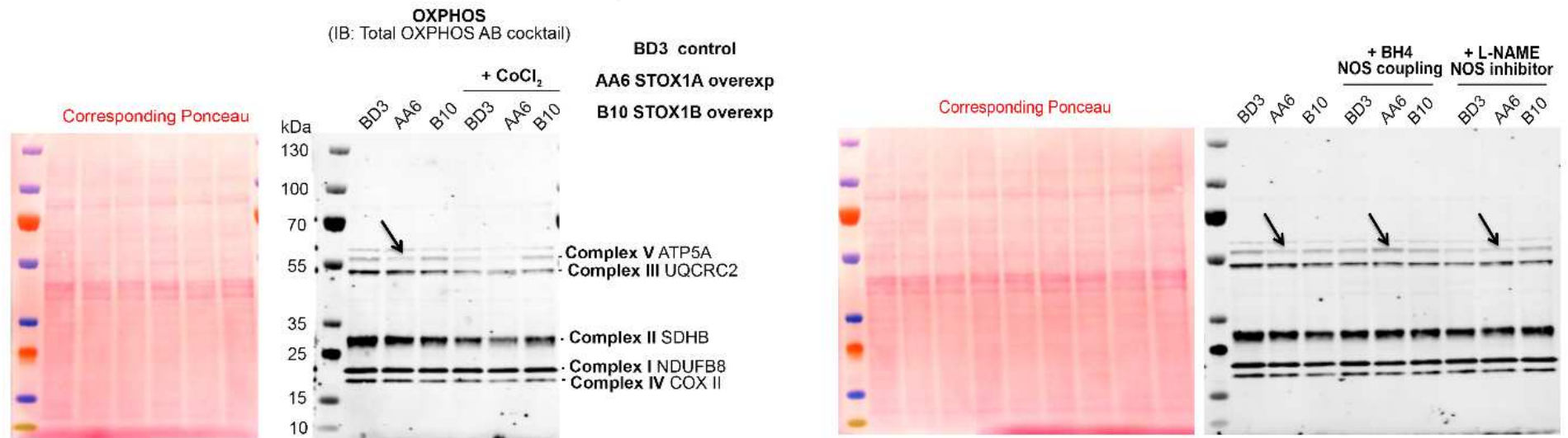
# Massive alterations of the placental transcriptome affects especially mitochondrial genes



(Doridot et al, Antioxidant and Redox Signaling, 2014)

# Specific alterations of mitochondrial complexes in *STOX1* overexpressing cells

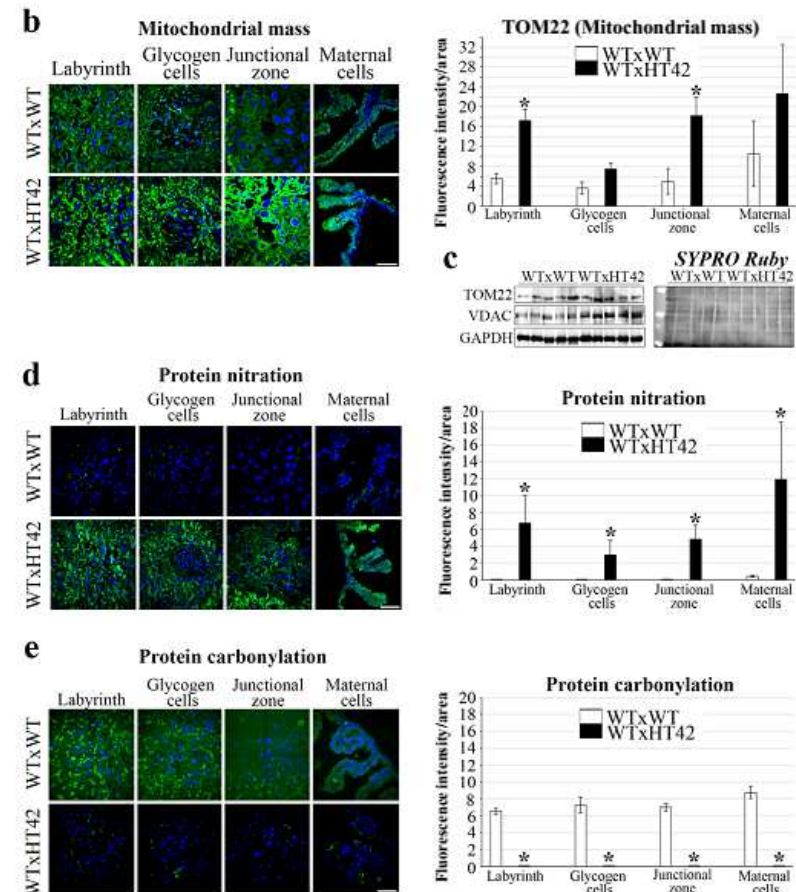
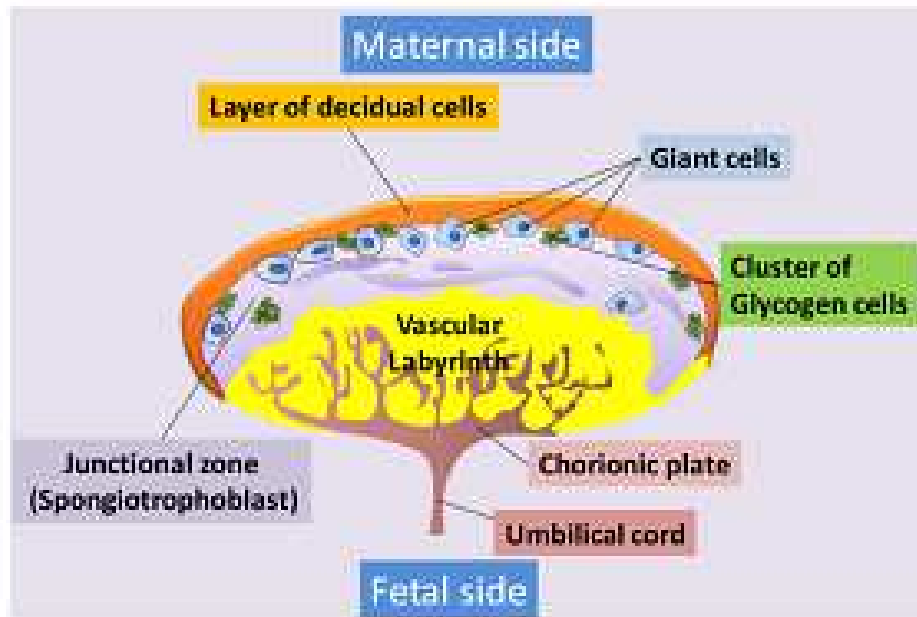
## STOX1A and STOX1B direct/indirect effect on OXPHOS complexes



- Complex I: no effect of STOX1A/STOX1B**
- Complex II: decrease due to STOX1A and STOX1B**
- Complex III: decrease due to STOX1A and STOX1B**
- Complex IV: decrease due to STOX1A and STOX1B**
- Complex V: decrease only due to STOX1A**

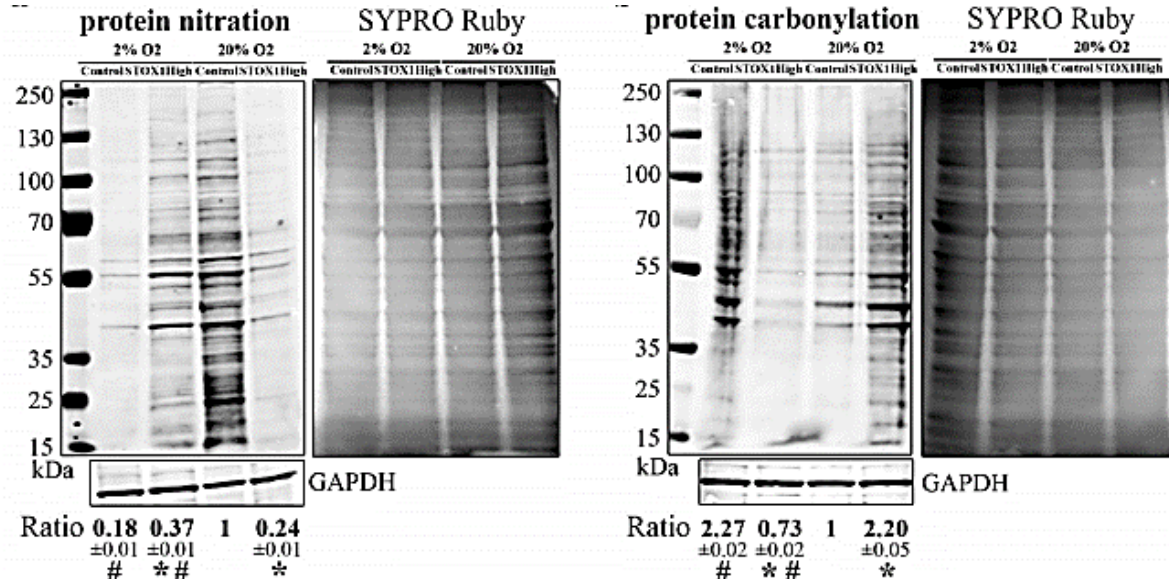
- BH4 treatment induces Complex V increase in AA6 (STOX1A overexp)**
- L-NAME treatment modifies the OXPHOS complexes in all cells types**

# At the placental level the oxidative/nitrosative stress balance is perturbed

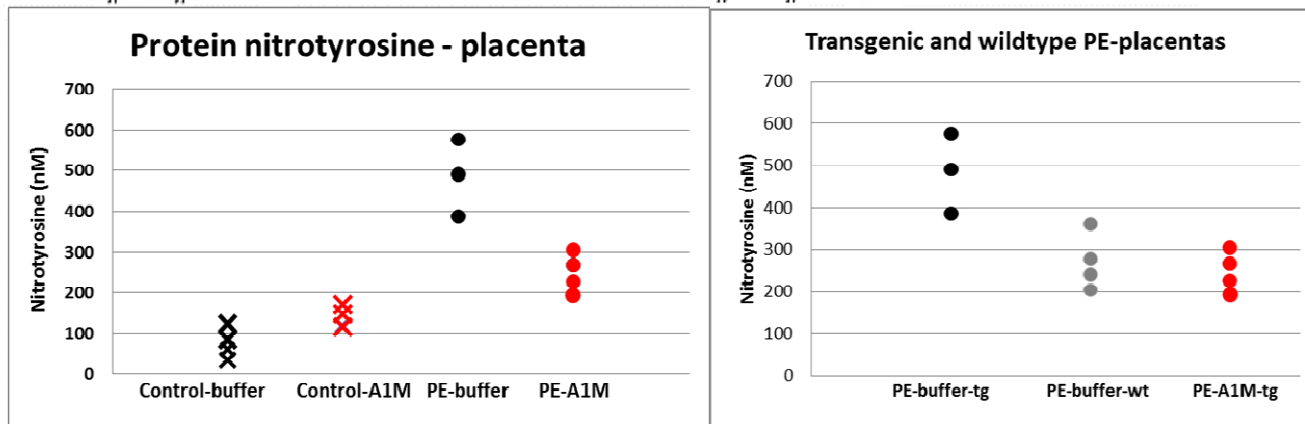




# Oxidative / Nitrosative stress : a mirror image in cells

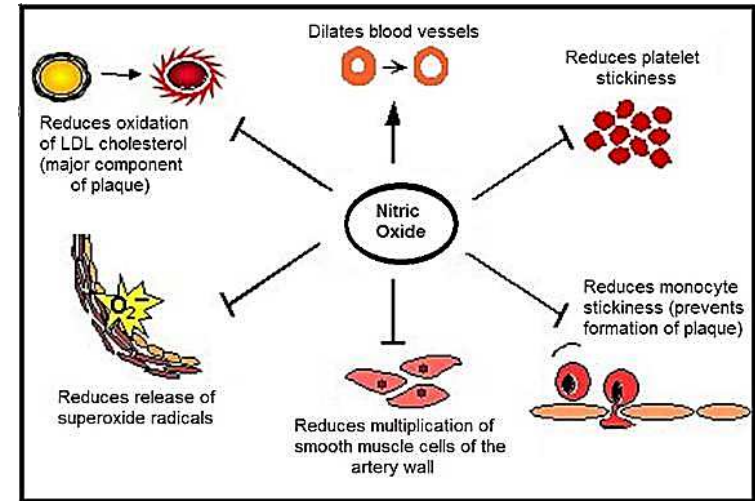
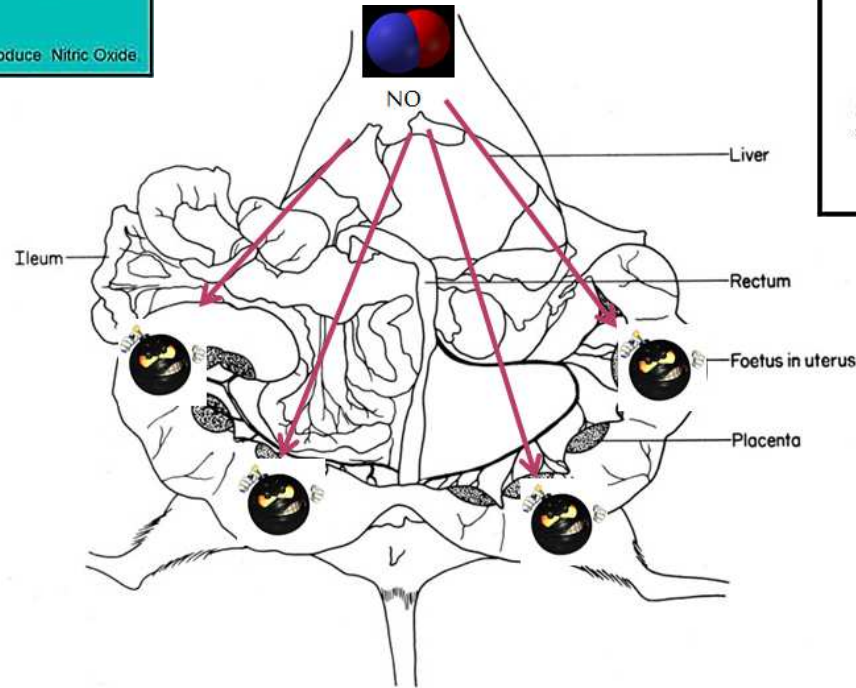
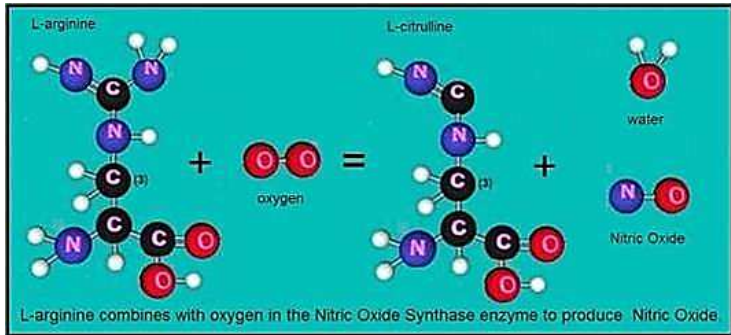


(Doridot et al, Antioxidant and Redox Signaling, 2014)



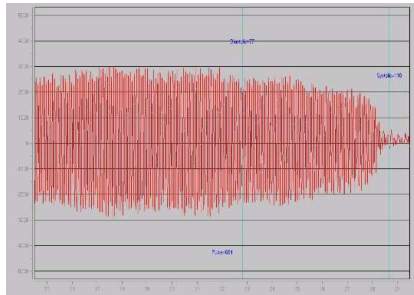
(Erlandsson et al, unpublished results)

# Pumping out NO in *STOX1* overexpressing placentas

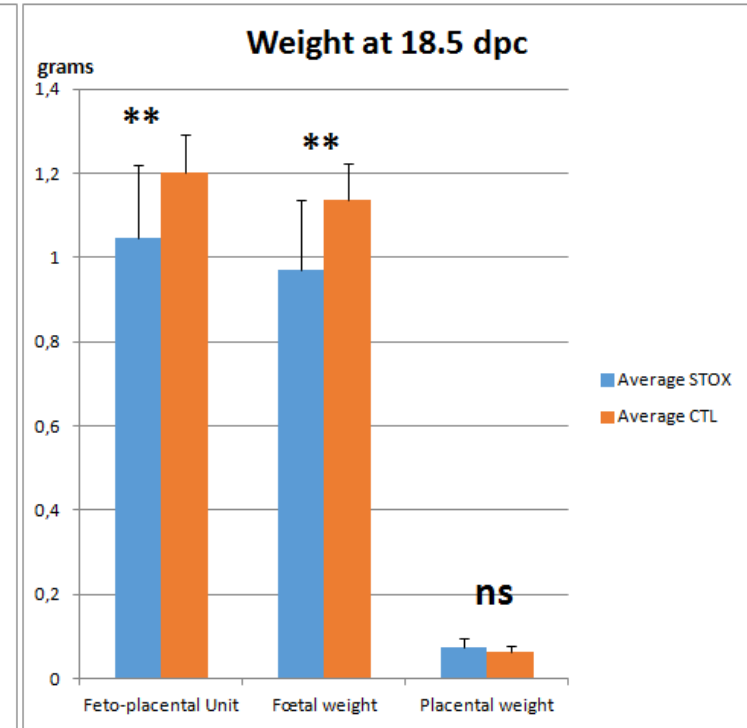
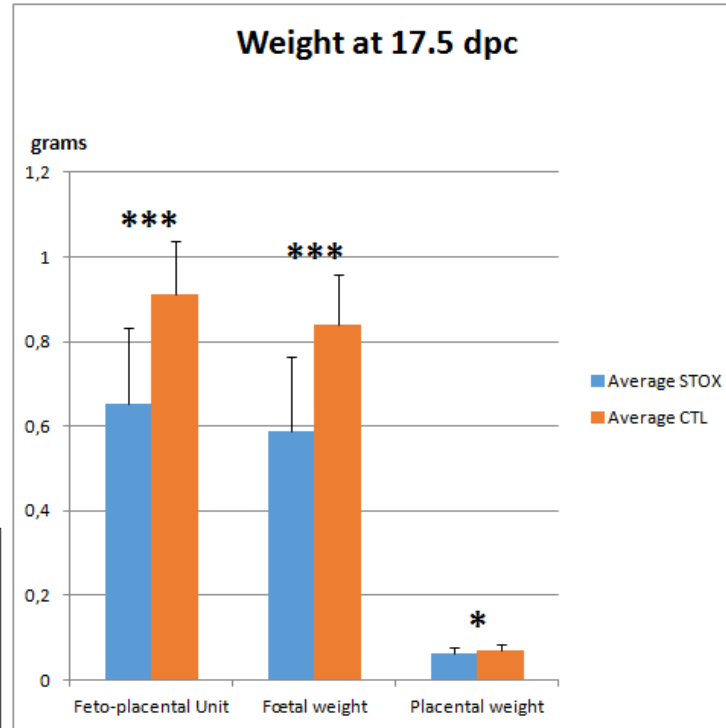
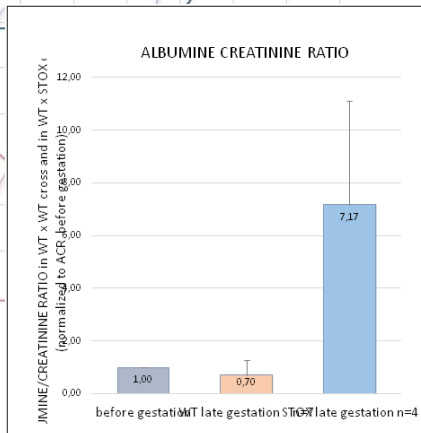
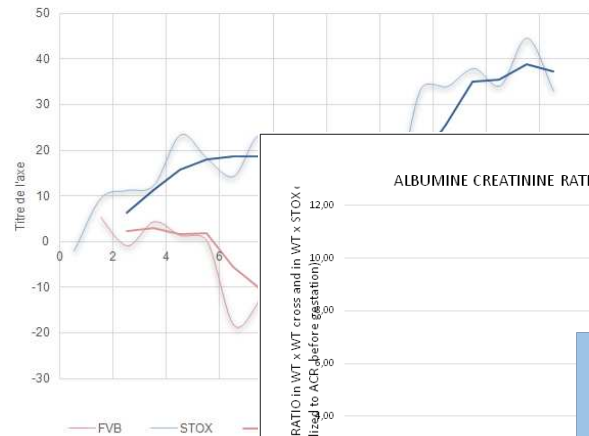


# Improved phenotyping: IUGR in the STOX1 mouse model

(Collinot et al, submitted)



Systolic Blood Pressure



Mesure de la pression par photopléthysmographie

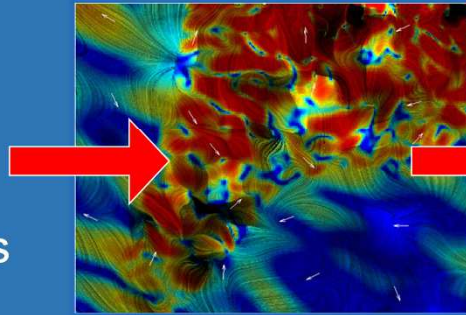


# BOLD(Blood-Oxygen-Level-Dependent) MRI analysis

What is the difference between **deoxyHb** and **oxyHb**?

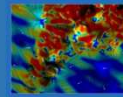
Remember T2\* and field inhomogeneities?

**DeoxyHb**  
paramagnetic  
strong field  
inhomogeneities



Fast dephasing  
Fast T2\*

**OxyHb** diamagnetic  
weak field  
inhomogeneities



Slower dephasing  
slower T2\*

# Specific values of the T2\* and $\Delta T2^*$ parameter

Organ	N	T2* ambient air period in msec (mean $\pm$ SD)	T2* hyperoxygenation in msec (mean $\pm$ SD)	P value
Maternal liver	27	11.9 $\pm$ 4.1	14.5 $\pm$ 5.2	<0.001
WT x TgSTOX1 mice				
Placental inner layer	83	14.9 $\pm$ 4.9	23.05 $\pm$ 8.2	<0.001
Placental outer layer	65	8.4 $\pm$ 3.3	14.5 $\pm$ 6.9	<0.001
Fetal brain	67	38.1 $\pm$ 8.0	43.4 $\pm$ 10.4	<0.001
Fetal liver	93	8.9 $\pm$ 1.8	11.3 $\pm$ 4.2	<0.001
WT x WT mice				
Placental inner layer	57	14.8 $\pm$ 4.1	20.7 $\pm$ 6.3	<0.001
Placental outer layer	52	8.2 $\pm$ 2.0	12.1 $\pm$ 3.6	<0.001
Fetal brain	39	41.4 $\pm$ 6.1	50.6 $\pm$ 9.9	<0.001
Fetal liver	53	9.5 $\pm$ 1.8	12.04 $\pm$ 3.0	<0.001

Mice	Placenta Inner Layer T2* ambient air period in msec Median (Q1-Q3)	Placenta Outer Layer T2* ambient air period in msec Median (Q1-Q3)	P value
WT x TgSTOX1 mice	15.2 (11.7-19.7)	7.9 (6.4-10.3)	<0.001
WT x WT mice	14.3 (11.9-16.9)	7.4 (6.1-9.8)	<0.001

Placental zone	WT x TgSTOX1 mice T2* ambient air period in msec Median (Q1-Q3)	WT x WT mice T2* ambient air period in msec Median (Q1-Q3)	P value
Placental inner layer (n=140)	15.2 (11.7-19.7) (n=83)	14.3 (11.9-16.9) (n=57)	0.924
Placental outer layer (n=117)	7.9 (6.4-10.3) (n=65)	7.4 (6.1-9.8) (n=52)	0.316

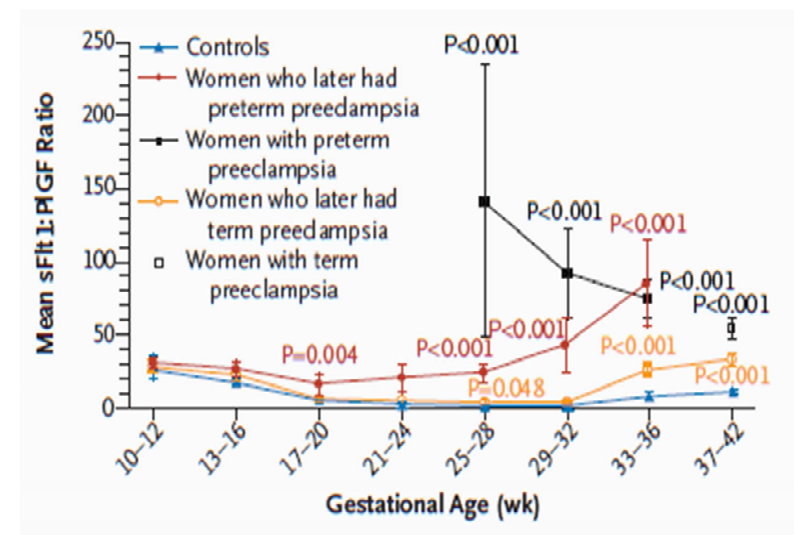
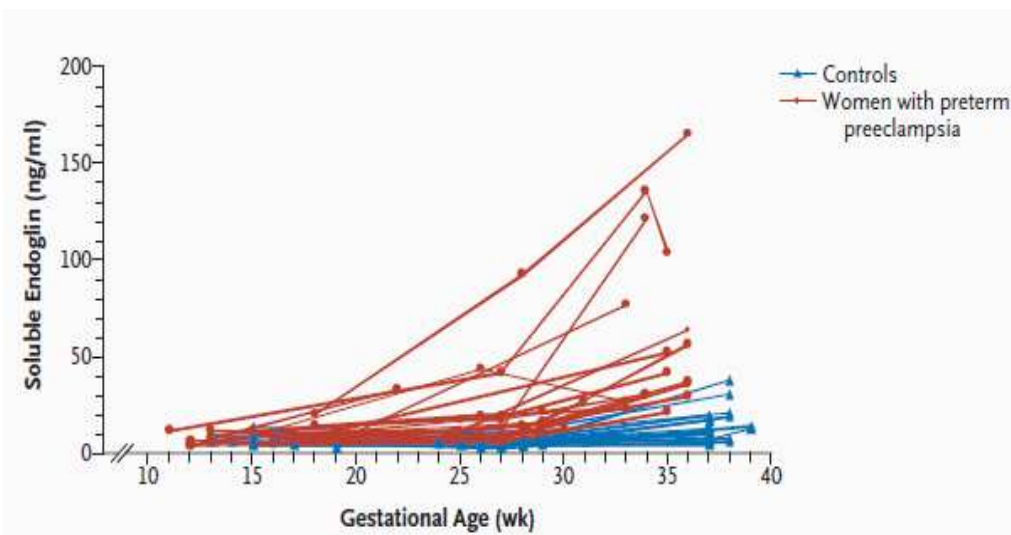
Placenta	WT x TgSTOX1 mice $\Delta T2^*$ (ms) (mean $\pm$ SD)	WT x WT mice $\Delta T2^*$ (ms) (mean $\pm$ SD)	P value
Inner layer (n = 140)	8.2 $\pm$ 5.9 (n=83)	5.8 $\pm$ 5.9 (n=57)	0.025
Outer layer (n=117)	6.1 $\pm$ 5.1 (n=65)	3.9 $\pm$ 3.1 (n=52)	0.005

Organ	WT x TgSTOX1 mice $\Delta T2^*$ (ms) (mean $\pm$ SD)	WT x WT mice $\Delta T2^*$ (ms) (mean $\pm$ SD)	P value
Fetal brain (n=106)	5.3 $\pm$ 6.1 (n=67)	9.1 $\pm$ 8.0 (n=39)	0.004
Fetal liver (n=140)	2.4 $\pm$ 3.7 (n=93)	2.5 $\pm$ 2.6 (n=53)	0.25

# Diagnosis and Prognosis of preeclampsia using markers

## Context

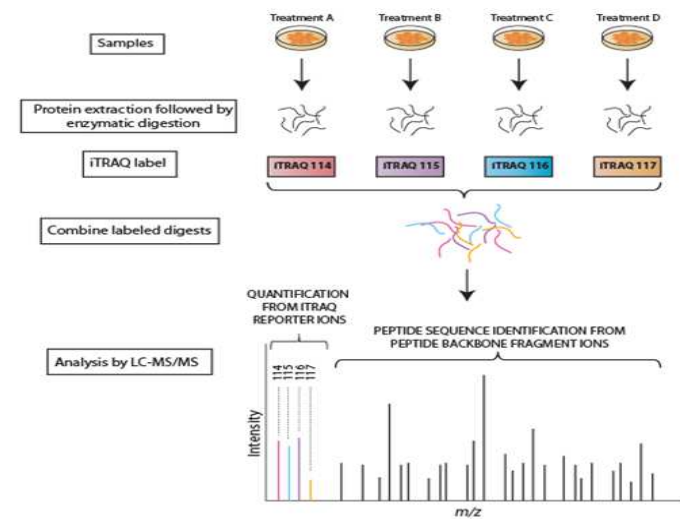
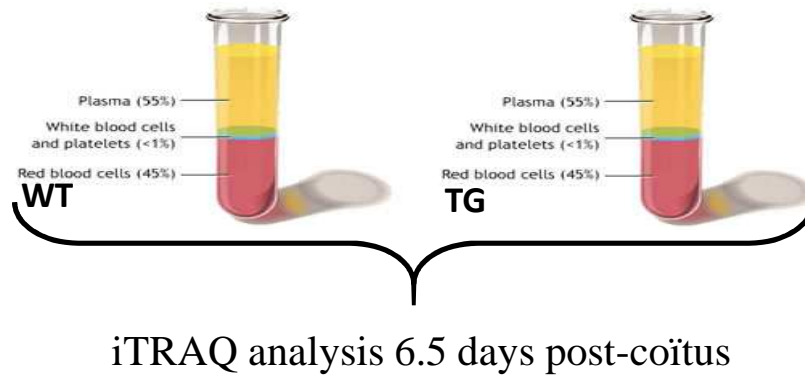
- Low-dose aspirin before the 16th week is efficient to protect from preeclampsia.
- Plasmatic presymptomatic markers of preeclampsia have been identified (sFLT1, sENG, PLGF, PAPPA); at best the difference is detected after the 20th week of pregnancy.
- Novel earlier marker are warranted.



Innovation à l'Institut Cochin; 16 juin 2017

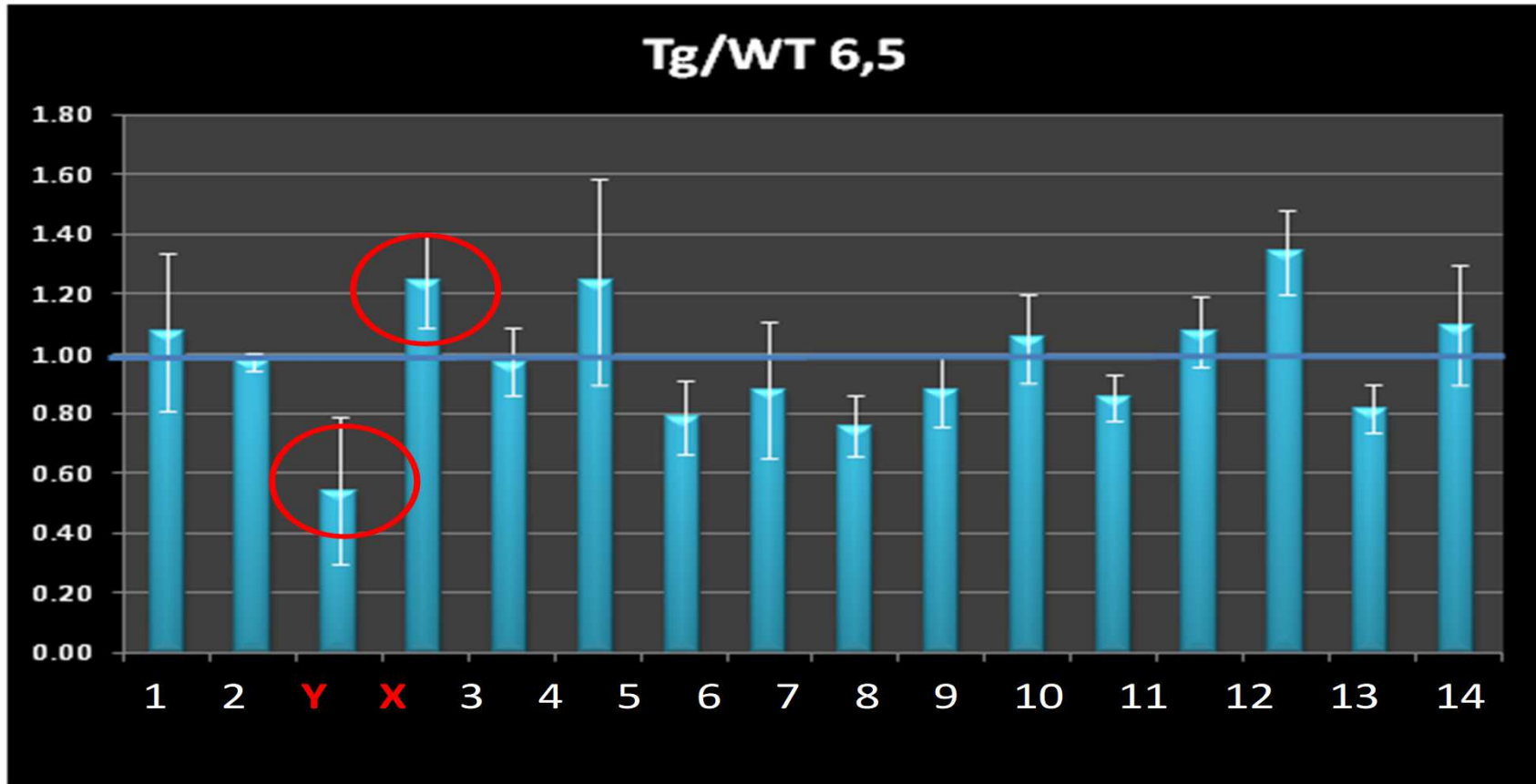
Levine et al, NEJM 2006

# Using the STOX1 mouse model to identify novel markers



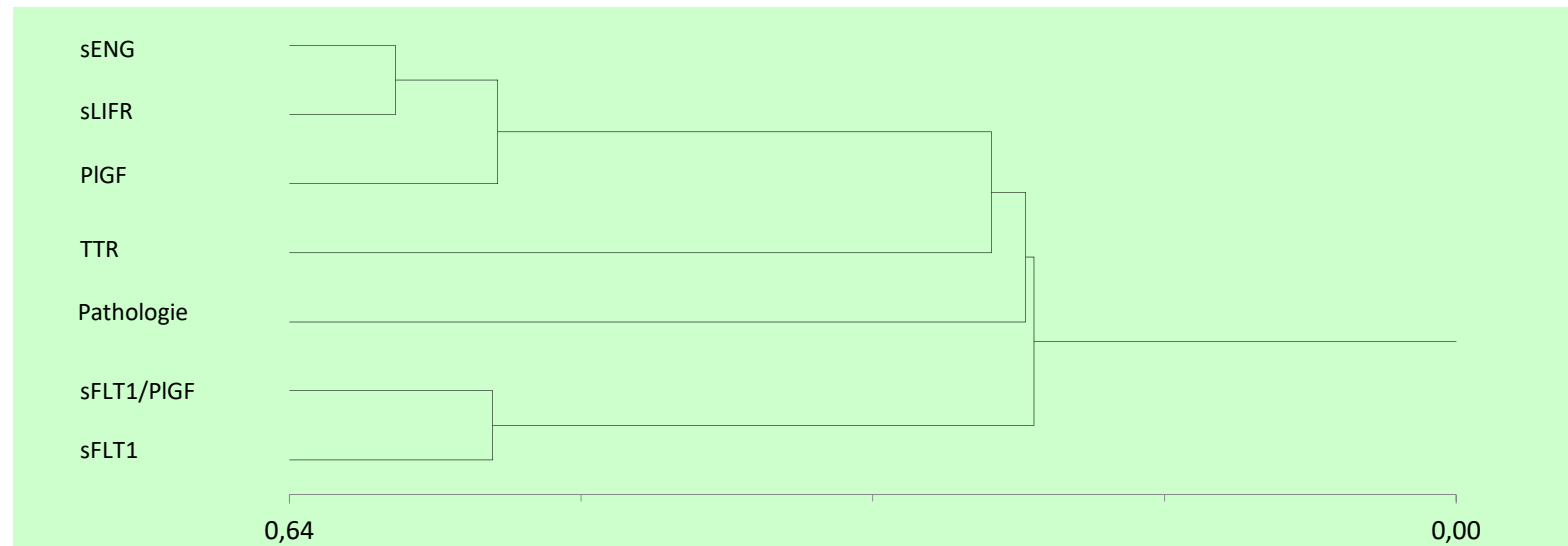


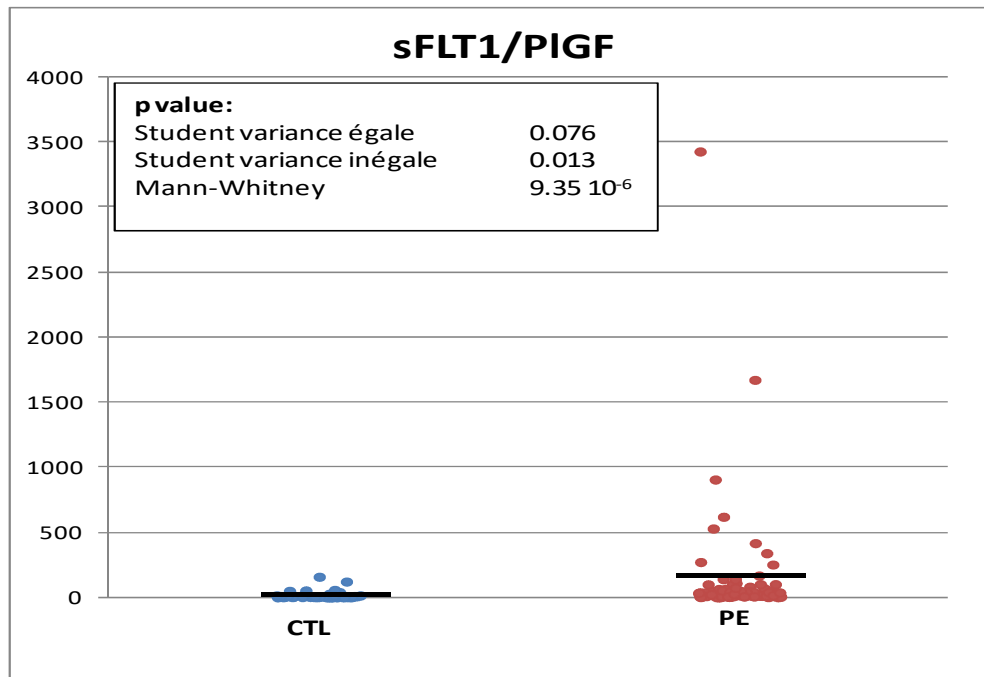
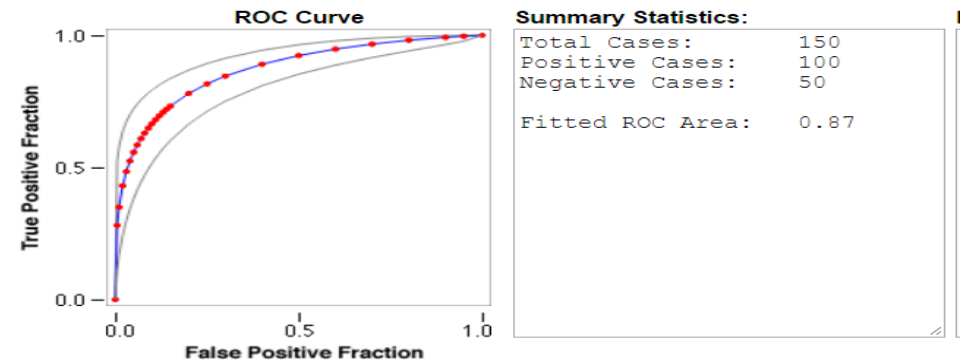
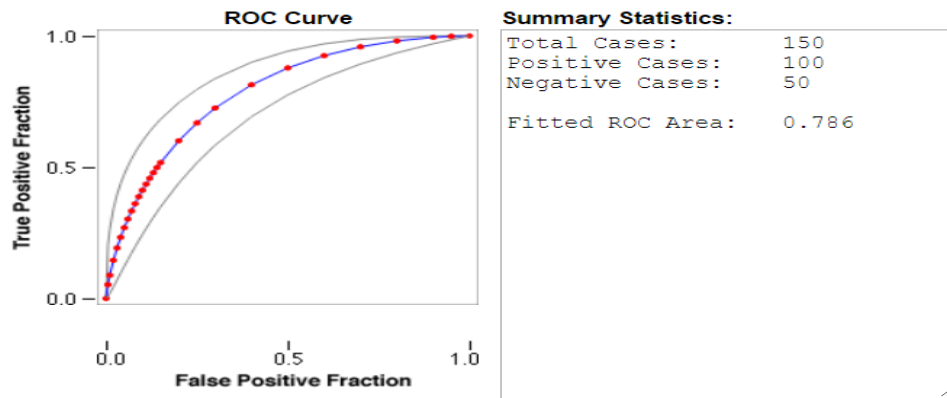
# iTRAQ results



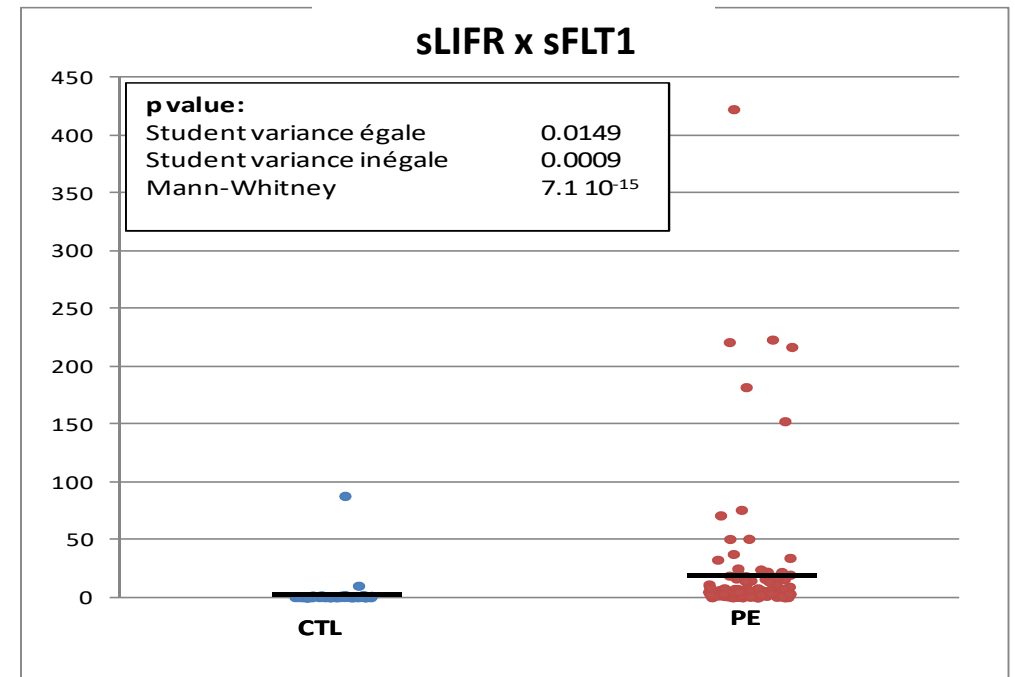
# Five markers were analyzed in 150 human plasma sample (Lund University)

Pearson Correlations and p-values							
Pathologie	-	0.232	-0.220	-0.153	0.236	0.078	0.146
sFLT1	0.004	-	0.047	0.003	0.105	0.147	0.528
TTR	0.007	0.568	-	0.255	0.017	0.221	0.065
PLGF	0.061	0.967	0.002	-	0.526	0.511	-0.097
sLIFR	0.004	0.200	0.839	4.99E-12	-	0.582	-0.062
sENG	0.340	0.073	0.007	2.42E-11	5.81E-15	-	0.128
sFLT1/PLGF	0.074	3.64E-12	0.426	0.238	0.454	0.119	-





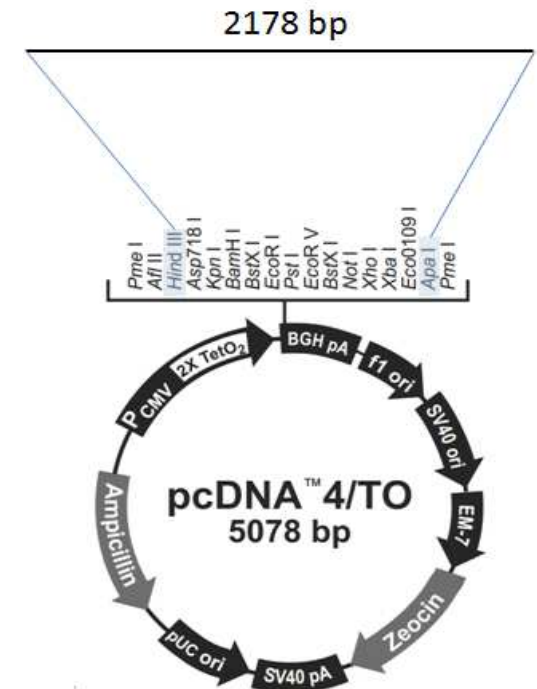
Mean: 14 vs 114



Mean: 2.4 vs 23.4

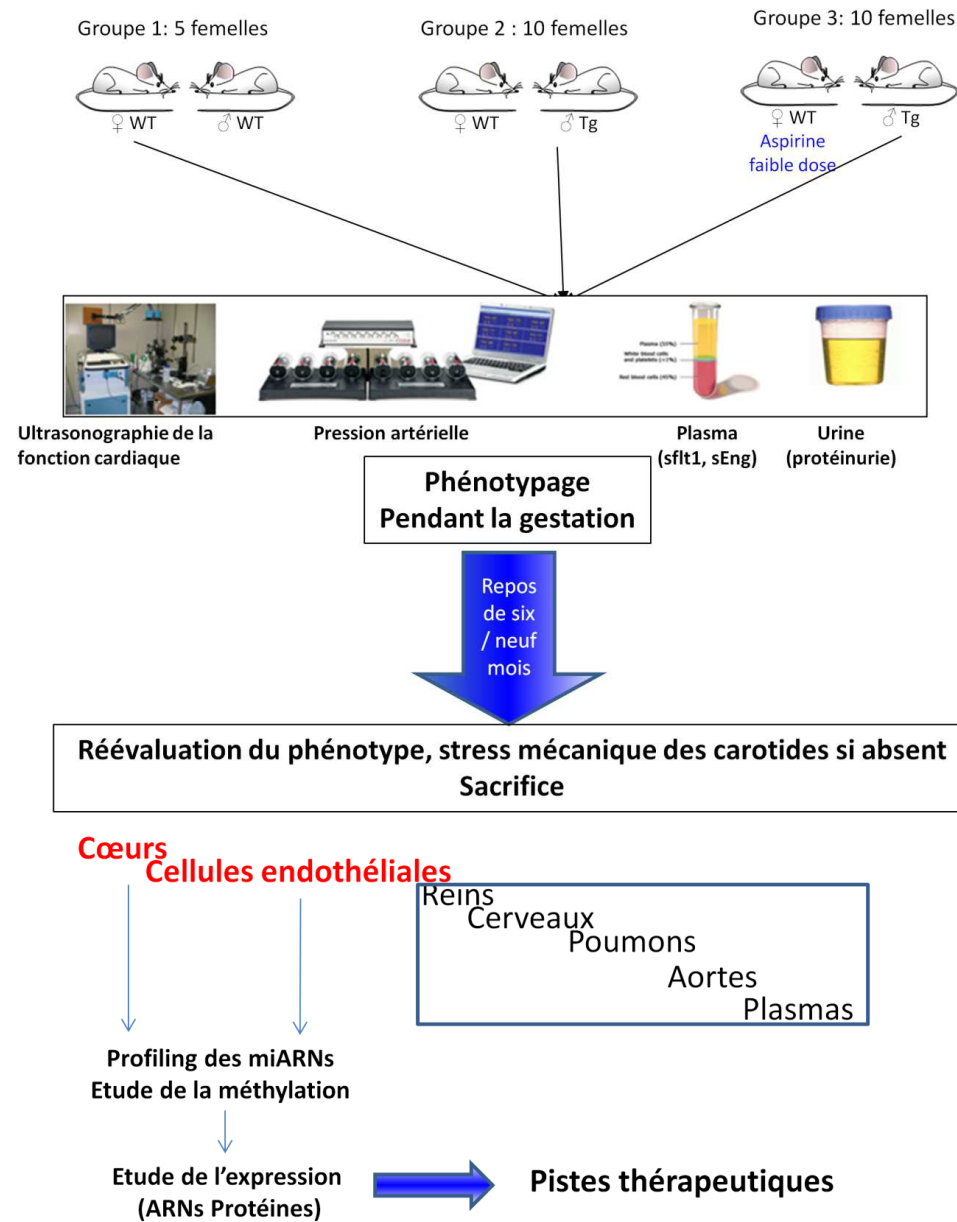
# Follow-up

1. Analysis of other additional human cohorts.
2. Generation of a Doxycyclin-inducible mouse model of overexpression of sLIFR ( )
3. Analysis of the phenotype (arterial hypertension/embryonic resorption, proteinuria) in function of the induction moment of the transgene

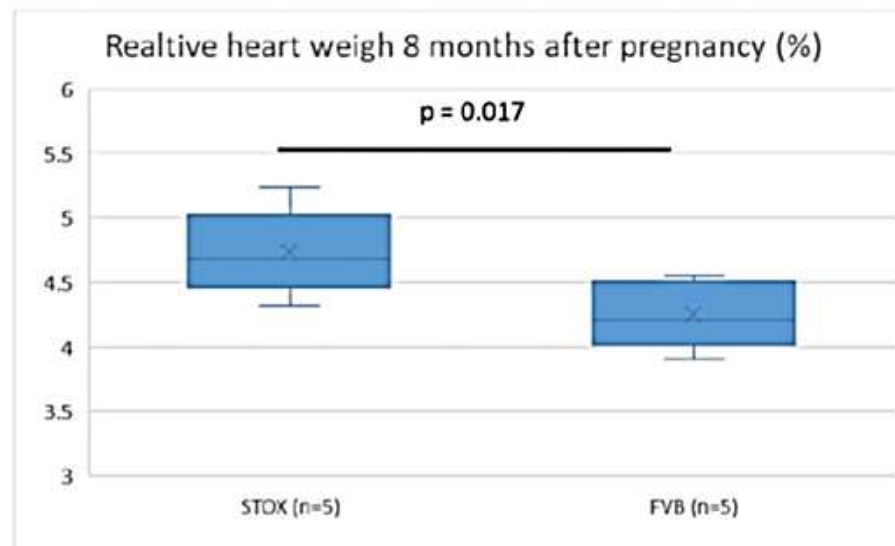




# Long-term effects of preeclampsia

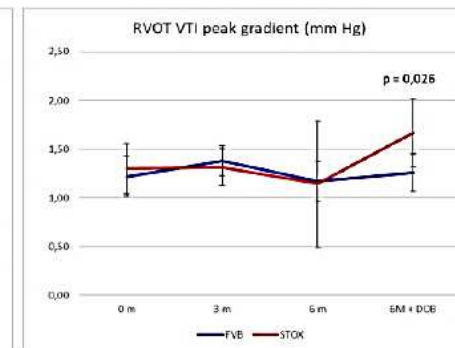
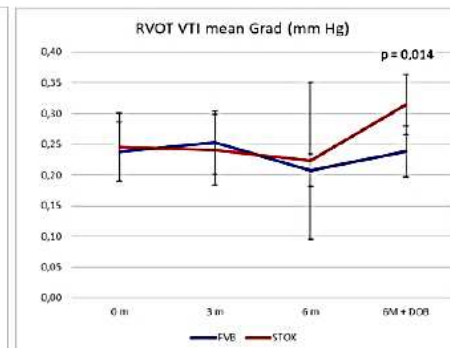
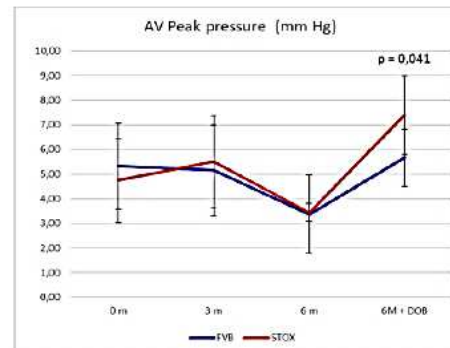
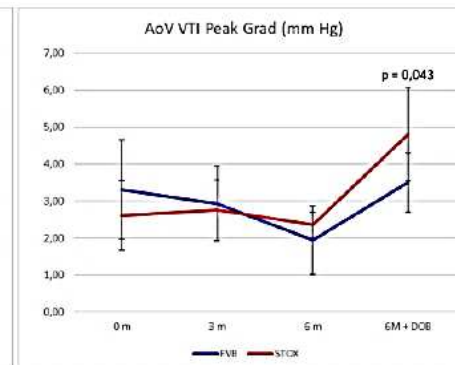
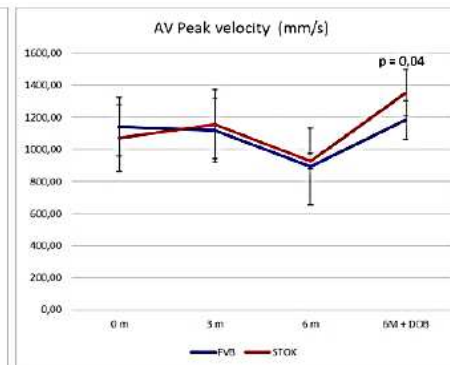
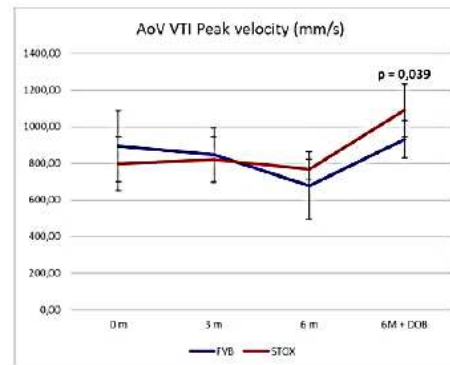
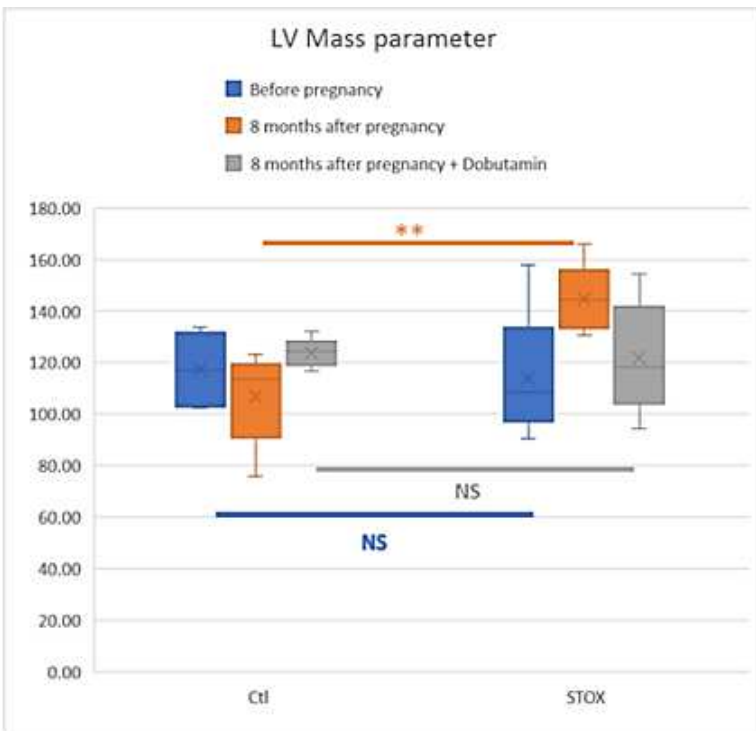


# Increased mouse heart weight 8 months after pregnancy after a preeclamptic pregnancy

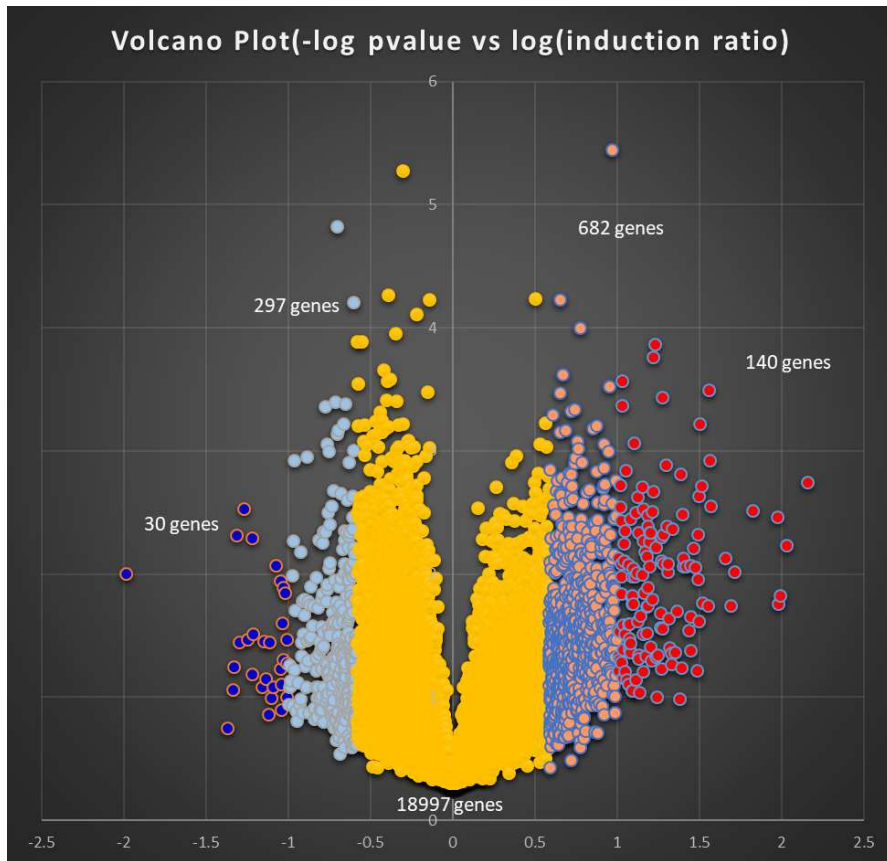


**Figure 1 :** Mouse heart weight is durably increased following a preeclamptic pregnancy. The difference is about 11% after correction for the mouse total weight. This difference was similar to the one observed at the end of pregnancy (Ducat et al, 2016) suggesting that the alterations were not reversible.

# An altered response to stress stimuli at the heart level (Doppler US analysis)



# Massive alteration of the endothelial cell transcriptome

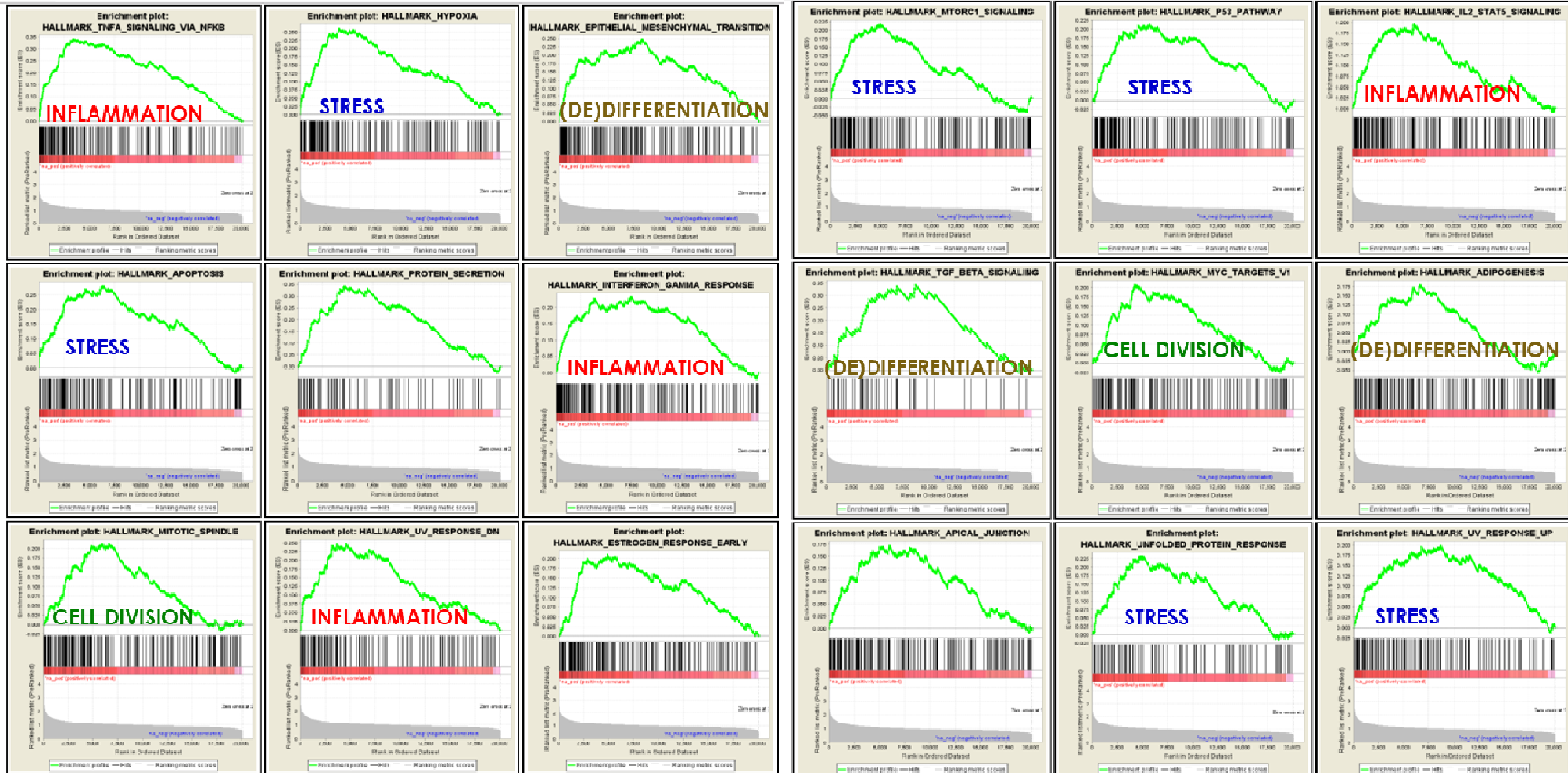


RIKEN cDNA 1190007107 gene  
 proteoglycan 2, bone marrow  
 CD27 antigen  
 cDNA sequence AB124611  
 ribosomal protein S27-like  
 deltex 3, E3 ubiquitin ligase  
 RIKEN cDNA 2900060B14 gene  
 developmentally regulated GTP binding protein 2  
 CD59a antigen  
 xeroderma pigmentosum, complementation group A  
 oxysterol binding protein-like 1A  
 lymphocyte cytosolic protein 1  
 RIKEN cDNA I830077J02 gene  
 DEAH (Asp-Glu-Ala-His) box polypeptide 8  
 intraflagellar transport 22  
 killer cell lectin-like receptor, subfamily D, member 1  
 thymocyte expressed, positive selection associated 1  
 protein tyrosine phosphatase, receptor type, C  
 RIKEN cDNA 1110059G10 gene  
 sarcospan  
 coiled-coil domain containing 6  
 TSSK6 activating co-chaperone  
 S100 calcium binding protein A8 (calgranulin A)  
 histone cluster 1, H2bc  
 tectonic family member 3  
 DEAH (Asp-Glu-Ala-His) box polypeptide 32  
 G protein-coupled receptor kinase-interactor 1  
 potassium voltage-gated channel, subfamily G, member 1  
 transmembrane protein 159  
 tryptophan hydroxylase 1

MKL/myocardin-like 2  
 muscle glycogen phosphorylase  
 actinin, alpha 1  
 glutathione S-transferase, mu 1  
 plexin domain containing 2  
 colony stimulating factor 2 receptor, beta 2, low-affinity (granulocyte-macrophage)  
 dynamin 3  
 peripheral myelin protein 22  
 src homology 2 domain-containing transforming protein C1  
 apolipoprotein D  
 sprouty homolog 4 (Drosophila)  
 developmentally regulated GTP binding protein 1  
 activating transcription factor 1  
 A kinase (PRKA) anchor protein (gravin) 12  
 insulin receptor  
 mitogen-activated protein kinase kinase kinase 3  
 decorin  
 TCDD-inducible poly(ADP-ribose) polymerase  
 thrombospondin 1  
 GTP binding protein (gene overexpressed in skeletal muscle)  
 thrombospondin, type I, domain containing 7A  
 voltage-dependent anion channel 2  
 matrix-remodelling associated 8  
 a disintegrin-like and metalloproteinase (reprolysin type) with thrombospondin type 1 motif, 4  
 protein phosphatase 3, regulatory subunit B, alpha isoform (calcineurin B, type I)  
 retinol saturase (all trans retinol 13,14 reductase)  
 RAS p21 protein activator 4  
 interleukin 6  
 ribosomal protein L7-like 1  
 RAB8B, member RAS oncogene family



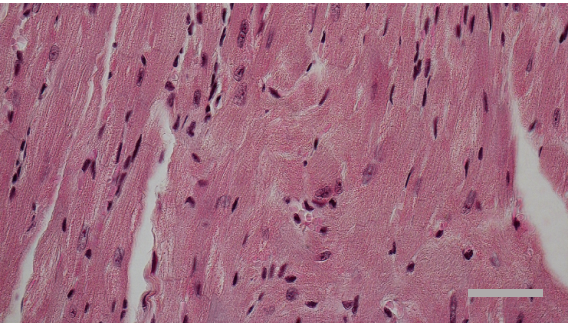
# Transcriptome functional clustering



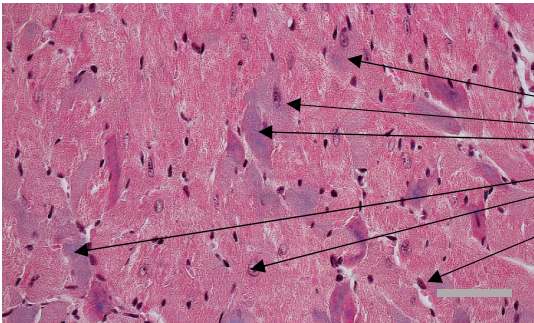
NES : 5.49 to 2.75

# Preliminary Results on Cardiac histology

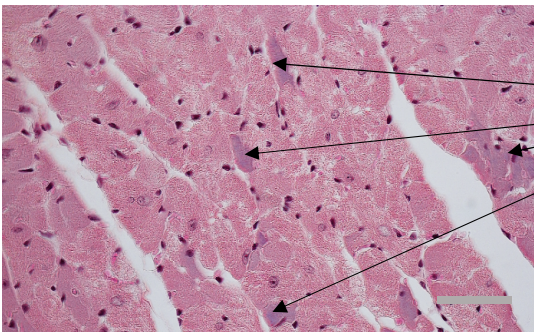
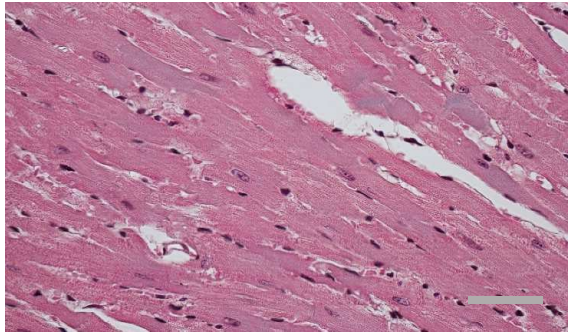
CTL pregnancy



STOX pregnancy



Fibrosis marks



Fibrosis marks

Trichrome Masson staining (x40, bar = 50µm)

## Institut Cochin

- **Cell and animal models**
  - Ludivine Doridot, PhD
  - Aurélien Ducat, PhD
  - Rosa Calicchio, PhD
  - Betty Couderc, Master Student
  - Irène Gaillard, Master Student
  - Sophia Palfray
- **Today**
  - Louise Biquard
  - Rajaa Aouache
  - Francisco Miralles
  - Céline Méhats
- **Analysis of the oxidative stress**
  - Christiane Chéreau
  - Frédéric Batteux

# Thank you for your attention

## Institut Pasteur (Mitochondrie et stress oxydatif)

- Laurent Châtre
- Miria Ricchiatti

## INRA (Jouy en Josas)

- Bruno Passet
- Johan Castille
- Marthe Vilotte
- Jean-Luc Vilotte

## Lund University

- Lena Erlandsson
- Stefan Hansson
- Grigorios Karampas
- Eva Hansson

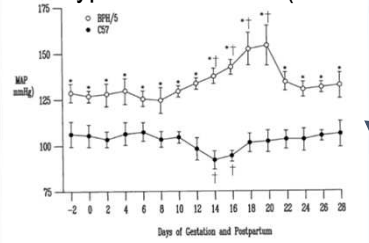
## Glasgow University

- Dylis Freeman
- Shahzya Huda

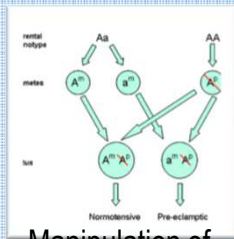


## Genetics

### Pre hypertensive mice (BPH/5)



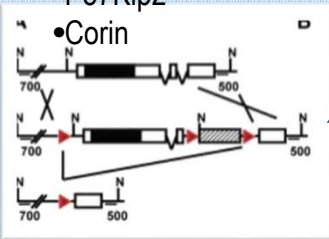
### Crosses (CBAXDBA)



Manipulation of imprinted genes

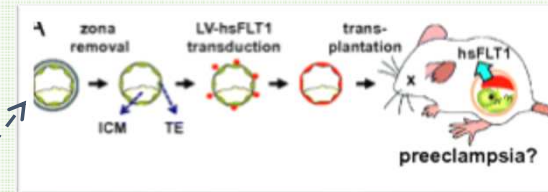
### Specific KO mice

- Comt
- P57Kip2
- Corin

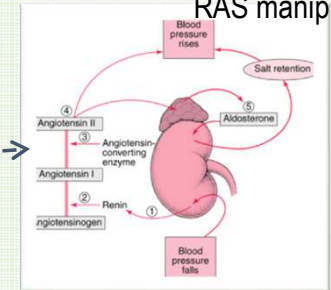


## Angiogenesis and Hypertension

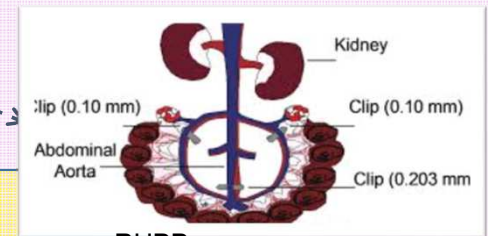
### Sflt1 overexpression



### RAS manipulation

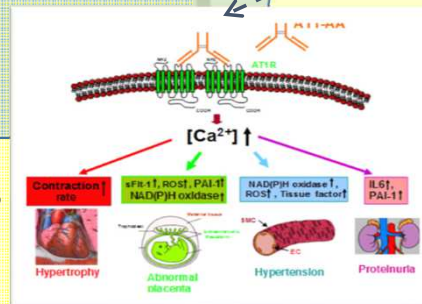


## Surgery

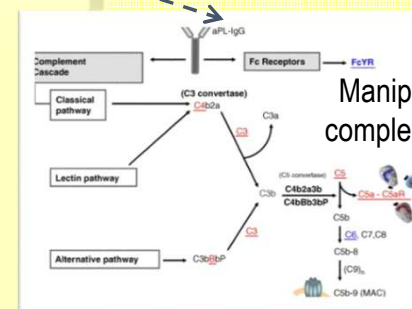


RUPP surgery

### Injection of autoantibodies



### Manipulation of the complement cascade



## Immunity and inflammation